The Contribution of Resident Vascular Stem Cells to Arterial Pathology

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Intimal accumulation of smooth muscle cells contributes to the development and progression of atherosclerotic lesions and restenosis following endovascular procedures. Arterial smooth muscle cells display heterogeneous phenotypes in both physiological and pathological conditions. In response to injury, dedifferentiated or synthetic smooth muscle cells proliferate and migrate from the tunica media into the intima. As a consequence, smooth muscle cells in vascular lesions show a prevalent dedifferentiated phenotype compared to the contractile appearance of normal media smooth muscle cells. The discovery of abundant stem antigen-expressing cells in vascular lesions also rarely detected in the tunica media of normal adult vessels stimulated a great scientific debate concerning the possibility that proliferating vascular wall-resident stem cells accumulate into the neointima and contribute to the progression of lesions. Although several experimental studies support this hypothesis, others researchers suggest a positive effect of stem cells on plaque stabilization. So, the real contribute of vascular wall-resident stem cells to pathological vascular remodelling needs further investigation. This review will examine the evidence and the contribution of vascular wall-resident stem cells to arterial pathobiology, in order to address future investigations as potential therapeutic target to prevent the progression of vascular diseases.

Keywords: Arterial wall, Intimal thickening, Smooth muscle cells, Resident stem cells, Smooth muscle cell heterogeneity

Introduction

Accumulation of smooth muscle cells (SMCs) in the tunica intima plays a crucial role in the pathogenesis of vascular lesions, including atherosclerosis and restenosis following angioplasty or stenting procedures (1, 2). In the classical pathogenetic hypothesis of atherosclerosis, follow-

ing injury growth factors and proteolytic agents induce SMC proliferation and migration from the tunica media into the intima (3). During this process, SMCs switch from a “contractile” to a “synthetic” phenotype, characterized from reduced expression of contractile proteins and enhanced response to growth and chemotactic factors (4). In atherosclerosis, SMC accumulation in the fibrous cap is monoclonal or oligoclonal (5, 6), implying that only a small number of medial SMCs proliferate in response to injury. This finding suggests the existence of a resident arterial stem cell subpopulation contributing to arterial healing in response to injury. Studies in human transplantation arteriopathy and primary atherosclerotic lesions showed that recruited bone marrow or host-derived circulating precursors are present in the lesions (6-8). Successively, other studies refused this hypothesis (9, 10), suggesting that SMCs in vascular lesions are mainly derived from the host parietal response. In parallel, the pres-

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ence of cells expressing stem cell antigens have been identified in the normal arterial wall (11, 12), supporting a potential role of adult vascular wall-resident stem cells (VRSCs) in vascular pathobiology. In this review, the characteristics and the potential contribution of VRSCs to the development of great vessel lesions are discussed.

Adult arterial vascular smooth muscle cells are phenotypically heterogeneous

Classically, SMCs within the normal tunica media of the adult vascular wall are heterogeneous, and a “contractile” and “synthetic” phenotype identified (13, 14). SMCs dis-

Fig. 1. phenotypic heterogeneity of adult vascular smooth muscle cells. Rat aortic normal media SMCs (left column) display with the classical “hill-and-valley” confluent grow pattern when cultured in plastic dishes, a more dendritic shape with a marked extracellular matrix remodelling when cultured in collagen gel and display abundant α -smooth muscle actin (α -sm actin)-positive stress fibers in immunofluorescence (rhodamine, bottom). In contrast, neointimal VSMCs obtained fifteen days after ballooning (right column) display a monolayered and epithelioid appearance (top), grow in Indian files with bipolar junctions in collagen gel and contain very low amount of α -smooth muscle actin (bottom).

playing a synthetic phenotype can be identified more frequently in pathologic arteries and have been characterized *in vitro* (Fig. 1). Synthetic SMCs obtained from intimal

Table 1. Stem antigens of adult vascular wall-resident stem cells

Stem antigen/synonym	Localization/type
CD133	Transmembrane glycoprotein
CD34	Transmembrane sialomucin protein
Flt-1 (VEGFR-1)	Transmembrane kinase receptor
KDR (VEGFR-2)	Transmembrane kinase receptor
c-kit (CD117)	Transmembrane tyrosine-protein kinase receptor
CD45	Protein tyrosine phosphatase
CD14	Cytoplasmic endotoxin receptor
sca-1	Transmembrane class III tyrosine kinase receptor
Notch-1	Transmembrane regulatory receptor

aortic tissue fifteen days after balloon injury displayed a monolayered epithelioid phenotype, with a cobblestone morphology, markedly different from the “hill-and-valley” growth pattern typical of normal medial VSMCs (14, 15). Moreover, neointimal cells express low amount of myocytic markers and differences are maintained in clonal expansion (14-16). It is worth of noting contractile and synthetic phenotypes are not permanent and can partially revert after stimulation with growth factors and extracellular matrix molecules (17-19). Phenotypically-regulated activation of proteins and receptors regulates differences in terms of proliferation and survival, being proliferating aortic neointimal VSMCs more sensitive to apoptosis (20). likely linked to a phenotype-regulated different NF- κ B activity (21, 22). Phenotype-dependent expression of integrins in vascular SMC regulates morphology, motility and gene expression in collagen matrix (Fig. 1; 18). A hy-

Fig. 2. Stem cell expression in rat aorta after injury and with aging. Serial immunostainings reveal (A) very rare cell are c-kit⁺ positive in normal 2 month-old rat aorta. (B) fifteen days after ballon injury, the majority of intimal and some of medial SMCs are c-kit⁺ positive. (C) Very rare SMCs are flt-1⁺ in normal 2 month-old rat aorta, but (D) the number of flt-1⁺ cells increases in 24-month old rat aorta.

pothesis to explain SMC heterogeneity in adult vessels is a different embryologic origin during vasculogenesis (23). In the chick embryo aorta, spindle-shaped and epithelioid phenotypes responding differently to TGF- β were isolated from distinct mesoderm and neural crest-derived regions (24). It is likely that the capacity of a different response to damage can be retained from SMCs during the adult life.

The existence of adult vascular wall-resident stem cells

Mesenchymal smooth muscle progenitors have been identified in the bone marrow, in the blood as circulating progenitors and in extravascular sites (25-27). Identification of these progenitor cells was mainly based on the finding of stem antigens shared from a SMC subpopulation in the normal arterial wall and/or in vascular lesions. Recent studies identified and characterised a small population of resident SMCs in the wall of great vessels of healthy adult mice expressing sca1 and low amounts of c-kit and CD34, absent in the adventitia (28). These putative VRSCs differ from bone marrow-derived smooth muscle progenitors or from those isolated from skeletal muscle, since they lack the ability to differentiate into lymphoid or myeloid lineages (29). A clonal subpopulation of vascular cells from the bovine aortic tunica media was phenotypically similar (CD29⁺, CD44⁺, CD14⁻, CD45⁻) to bone marrow-derived mesenchymal stem cells (30). Progenitor cells named "mesoangioblasts" isolated from explants of murine dorsal aorta display differentiative potential into various mesenchymal cell types other than myocytic cells *in vitro* and express both myogenic and endothelial markers (31, 32). The expression of main stem markers of adult VRSCs are listed in Table 1. Immunohistochemical investigation revealed c-kit⁺ and CD133⁺ cells in human atherosclerotic plaques and restenosis lesions and in rat aortic post-injury neointima, whereas SMCs in the tunica media of primary atherosclerotic plaques and normal arteries were c-kit⁻ (11, 33). Progenitor cells expressing sca-1, CD34, KDR and c-kit were detected within human plaques and the adventitia (11). Most of the knowledge concerning stem phenotype derives from studies of animal models of vascular injury. VEGFR-1 or flt-1⁺ and c-kit⁺ SMCs are extremely rare in normal rat aorta, whereas their number dramatically increases fifteen days after injury (Fig. 2), with many of intimal cells co-expressing both α -smooth actin and flt-1 or c-kit (12). After sixty days, after complete re-endothelialization stem marker-expressing intimal cells almost disappeared and α -smooth actin expression restored (12). Double immunostaining experiments revealed that flt-1⁺

Fig. 3. flt-1 expression and smooth muscle cell proliferation. Double immunostaining reveals that flt-1 expression (red) characterizes rat aortic bromodeoxyuridine+ proliferating SMC (arrow, black) in the tunica media three days after ballooning.

medial are also positive for anti-bromodeoxyuridine (Fig. 3) or apoptotic TUNEL staining, confirming the preferential involvement of flt-1⁺ cells in proliferative and apoptotic behaviour (22, 34, 35). Similarly, many intimal cells of mice carotid arteries express Hedgehog-induced Notch 1 early after injury (36).

The adventitial origin of vascular-wall resident stem cells

Although recent studies supported the hypothesis that the majority of SMCs in vascular lesions originates from the vessel wall, an open question remains the layer of origin of VRSCs. After early reports documenting that activation of adventitial fibroblasts precedes post-damage experimental neointimal proliferation (37), increasing evidence accumulated supporting the involvement of adventitial vascular progenitor cells in the development of arteriosclerosis, including transplant arteriosclerosis, angioplasty-induced restenosis, vein graft atherosclerosis, and spontaneous atherosclerosis. The adventitia of the arterial wall contains progenitor cells, which can differentiate into vascular SMCs. In apoE-deficient mice These progenitor cells were able to migrate from the adventitia into the intima, where they accumulate to contribute to atherosclerotic lesions of vein grafts (38). Hu et al. (2004) showed documented aortic adventitial cells in apoE-deficient mice expressing sca-1 and c-kit stem cell markers; moreover, ex vivo PDGFB-stimulated adventitial stem cell-expressing cells displayed a myocytic phenotype with the expression multiple smooth muscle markers (39). Lineage tracking

with bone marrow transplantation from ROSA 26 LacZ gene-expressing transgenic mice documented that adventitia-derived smooth muscle precursors migrate to the neointima in a vein graft atherosclerosis model, with no evidence of a bone marrow origin (39). The same Authors suggested that the extent of injury is crucial to regulate mobilization of adventitial progenitor cell and their contribution to vascular lesions. Several issues remain unresolved as to the physiologic relevance of adventitial progenitor cells, and if these cells are also documented in human lesions. Adult vascular adventitial tissue contains mature adipose tissue. Recently, an adipose-derived stem population has been well characterized (40–45). Adipose derived stem cells are capable to regenerate *de novo* mesenchymal tissue and are widely used for their pluripotential regenerative potential (46–52). In most of these studies, vascular transdifferentiation is reported as a crucial event to explain clinical efficacy of adipose-derived stem cells to ameliorate grafting and tissue regeneration. It is interesting to note that, differently from adipose-derived stem cells, bone marrow-derived circulating progenitor cells do not transdifferentiate into adipocytes *in vivo* and play little, if any, role in expanding the number of resident adipocytes during tissue growth (53). These findings strongly support that *de novo* vessel formation and/or vascular remodelling involve transdifferentiation of adventitial adipose-derived stem cells. Conversely, in long-term cultures adipose tissue-derived perivascular cells retained myogenicity and mesenchymal stem cell markers expression, similar to native, non-cultured perivascular cells (53). The vasculogenesis-promoting potential of adventitial adipose-derived stem cells depends on interaction with resident endothelial cells and involving contact and bi-directional interaction, resulting in modulated secretion of cytokines and extracellular matrix molecules, so leading to vascular remodeling (54). Cellular communication occurring between adventitial adipose-derived stem cells and endothelial precursor cells and triggered by cell-cell contact is partially mediated by secreted VEGF, but this effect is not observed by using HUVECs, suggesting that adipose-derived stem cells are unique in this sense (55). Being adventitial precursors a potential source of SMCs, the modulation of the contribution of these progenitor cells is a potential tool to counteract pathological vascular remodeling by cellular, genetic, and tissue engineering approaches.

The pericyte-like phenotype of adult vascular wall-resident stem cells

Blood vessel walls harbor a reserve of progenitor cells

that may be integral to the origin of non-vascular tissues as other related adult stem cells (56). Recently, resident pericyte-like cells from the adventitia have been hypothesized to contribute to restenosis. Rare NG2, PDGFR β and CD146-expressing cells identified in the adventitia of uninjured mouse femoral arteries in response to injury increased in number and expressed a SMC-like phenotype (31). Bone marrow transplantation and *ex vivo* artery culture strongly supported that pericyte-like progenitor cells are not bone marrow-derived, but originated from the adventitia after proliferation of resident perivascular pericyte-like cells (32). Cells with progenitor pericyte-like properties have been also isolated from the tunica media of adult rat aortas and are capable to generate spheroidal colonies in suspension and, when serum-cultured, retain CD34 negativity and express *de novo* SMC markers (32). The relationship between this population of resident adventitial pericyte-like cells, reparative myofibroblasts, normal media SMCs and other progenitor populations during pathological vascular remodelling is still matter of debate and need further investigation.

The contribution of adult vascular wall-resident stem cells to vascular lesion progression

The discovery of the existence of VRSCs has challenged the classical pathogenetic model of tunica media as unique source of SMCs contributing to vascular lesions (1, 2). Atherosclerosis is a chronic inflammatory disease, in which risk factors result in dysfunction and damage to the arterial endothelium and slow progression, characterized from intimal accumulation of SMCs in a fibromuscular cap covering a cholesterol-rich necrotic core (1). Both SMCs and monocyte-macrophages proliferate, accumulate and undergo apoptosis into the neointima (1, 2, 34, 35). In restenotic lesions, after acute injury, rapid proliferation and subsequent accumulation of SMCs from the tunica media into the intima results in rapid lumen occlusion. In these lesions, SMC progenitors have been identified by the expression of bone marrow or progenitor cell markers *in vivo*. Green fluorescent bone marrow transplantation in mice revealed that bone marrow-derived cells contribute up to 60% of SMCs in the post-injury neointima (7). Other studies report that donor-derived cells represent up to 10% of SMCs within atherosclerotic lesions in humans receiving sex-mismatched bone marrow transplantation (8). The major limit to these investigations was the method to estimate stem marker expression in lesional SMCs and accuracy of co-localisation of two markers, being confocal microscopy rarely performed. When accurate techni-

ques were applied, even in transplant arteriopathy studies, neointimal SMCs appeared to originate mainly from the recipient aortic wall (6, 9, 10). Moreover, other studies report that expression of stem markers is present only in early post-injury lesions, questioning about the transient nature of stem phenotype and the survival of these cells (22). Moreover, many studies employed α -smooth muscle actin as myocytic marker, but the latter is generally down-regulated in SMCs of vascular lesions (13), but strongly expressed in myofibroblast reparative cells (16). Consequently, those recent studies strongly support the hypothesis that the majority of SMCs in advanced atherosclerotic lesions and healing after plaque rupture originates from the vessel wall. Nevertheless, enhanced adventitial-derived progenitor cell recruitment and increased neointima formation has been seen after growth factor stimulation in animal models of vascular injury (12, 37, 38), so favoring vascular lesion progression. More recently, it has been reported that smooth muscle progenitor cell enrichment reduces the progression of early but not advanced atherosclerotic lesions plaques in mouse aortas, suggesting a favourable effect on plaque stabilization (57). These findings support a beneficial role of smooth muscle progenitor cells in suppressing atherosclerosis and its clinical sequelae, but still their parietal or extraparietal origin remain under investigation (58). More accurate studies demonstrating whether VRSCs either protect or promote vessel from pathological remodelling are needed, in order to promote cell-based or pharmacological approaches aimed to prevent vascular diseases (59).

Aging and vascular stem cell marker expression

Accumulation of vascular smooth muscle cells in the tunica intima plays a major role in the pathogenesis of atherosclerosis. Aging represents a risk factor for the development of vascular lesions (60-63) and the prevalence of atherosclerotic lesions in aged subject must be considered in therapeutic and experimental approaches (19, 64). Beside proliferation and differentiation, intrinsic aging enhances stem antigen expression in aortic SMCs (65) and, in parallel, susceptibility to atherosclerosis (60). Atherosclerotic lesions and also macroscopically normal human and rat aortas show an increased number of VEGFR-1⁺, c-kit⁺ and sortilin⁺ cells in the intimal thickening (65, 66). Both circulating and resident progenitor cells have been evocated to contribute to the response of the adult arterial wall to damage during atherogenetic process and aging (58). Inflammatory cell recruitment induces VEGFR-1-mediated physiological and pathological

angiogenesis that favors the increase of vessel lumen increase and its stabilization and counteracts pathological angiogenesis stimulated from PlGF-mutated variants that not bind VEGFR-1 (67, 68). These findings suggest that with aging stem cells with a synthetic VEGFR-1⁺ myocytic phenotype prevail and contribute to aortic myointimal thickening and to vascular angiogenetic/arteriogenetic and healing processes (69, 70). If this population derives from perpetual proliferation of VRSCs or phenotypic conversion of previously differentiated SMCs is still object of debate.

Conclusions

SMCs within the normal tunica media of the adult vascular wall are heterogeneous, and prevalence of a “synthetic” phenotype is found in atherosclerotic and restenotic lesions. Starting from SMC heterogeneity, a considerable accumulating evidence suggests the presence of a small population of VRSCs in different layers of the normal adult arterial wall, and its potential contribute to the homeostasis of post-natal arterial wall but also to pathological remodelling. Progenitor cell mobilisation may be tracked using genetic markers or co-localisation of stem and myocytic markers, but still variability of results is encountered according to the different experimental procedures and techniques. Further accurate studies are needed to define the positive or negative role of VRSCs in arterial pathobiology and to identify if the selective control of VRSC intraparietal mobilization may represent an attractive therapeutic opportunity.

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Potential conflict of interest

The authors have no conflicting financial interest.

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